

Personalised Medicine and Antibiotics: Should your DNA decide your dose?

Introduction

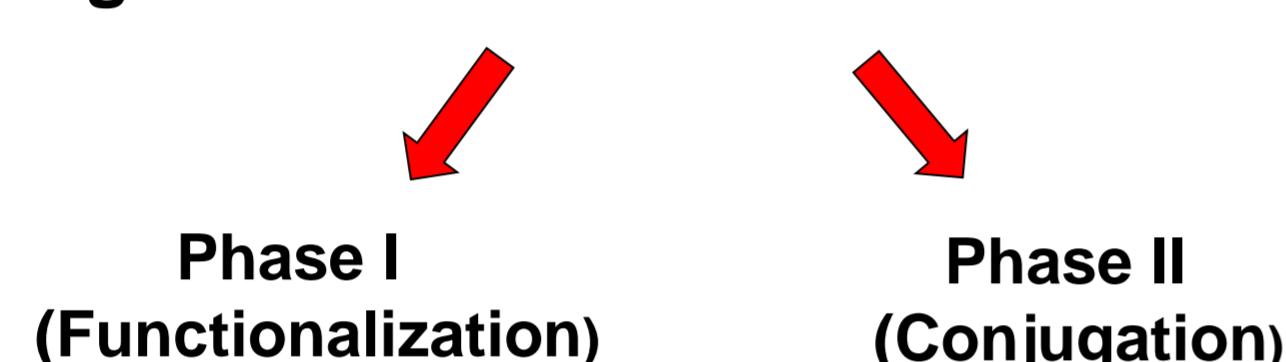
- Antibiotics remain the major treatment for bacterial infections
- Studies with other drugs show they are metabolised at different rates in different individuals, does this also apply to antibiotics??

Rapid metabolism of antibiotic = efficacy of the drug could be compromised and treatment could fail

OR

Slow metabolism of antibiotic = drug level remains high and increases likelihood of toxicity

Drug Biotransformation Reactions in Humans



- The enzymes that perform these reactions are primarily found in the liver and are highly polymorphic, meaning some people have non functional versions of these enzymes.

Testing Antibiotic Metabolism in Human Hepatoma cells - Method

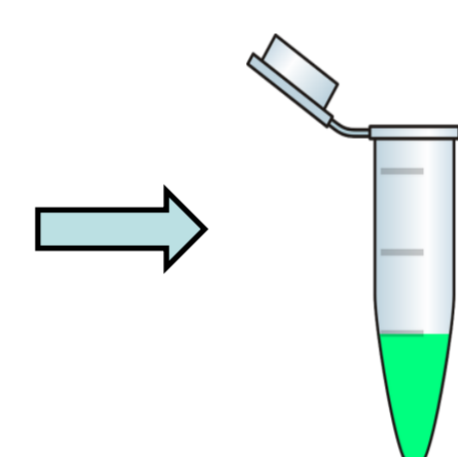


Liver cells seeded into 24 well plate at 1×10^5 cells per well

Cells left to attach and settle for 36 hrs



Cells washed with PBS and then given a $2\mu\text{g/ml}$ dose of Amoxicillin



Supernatant removed at Time 0 and 24hrs

Supernatant diluted (doubling dilutions to x8) - 100 μl in 96 well plate



100 μl of *Staph. Aureus* added to each well (1×10^5 /well)
Incubated at 37° C for 24 hrs (static)
Read OD 595



- Increased OD 595 = Reduction in level of active antibiotic present so allowing bacterial growth = liver cells have metabolised the antibiotic

Amoxicillin Metabolism by Human Hepatoma Cell Lines - Results

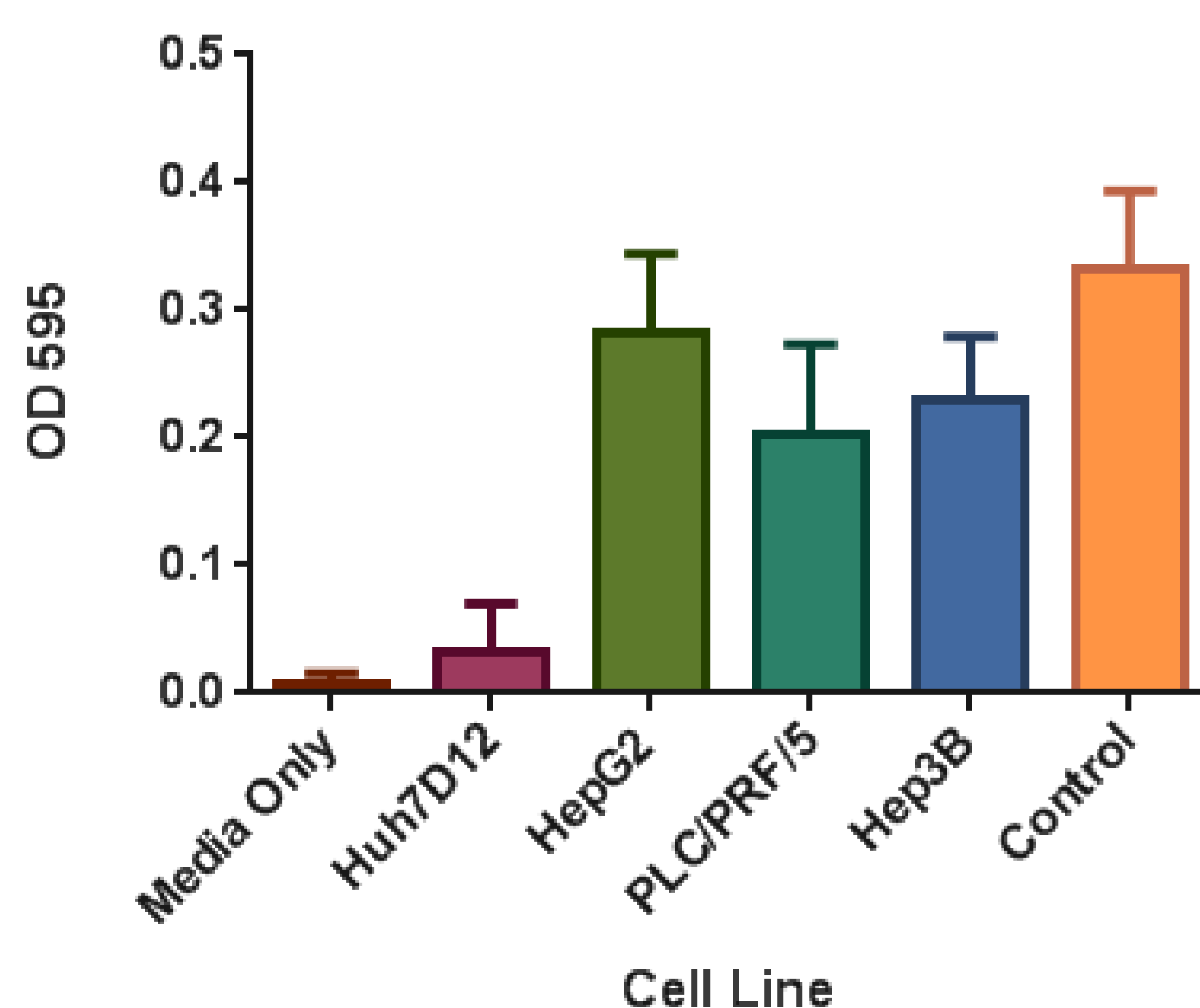


Figure 2. Amoxicillin metabolism by Human hepatoma cell lines. Media Only control = media plus antibiotic only, no liver cells present, confirms that amoxicillin does not naturally degrade after 24hrs at 37° C. Control = *Staph. Aureus* growth control, no antibiotic present. Data shown: 24hrs supernatant sample, x8 dilution.

Project Aim

Using Human Liver cancer cell lines:

- Map polymorphisms in genes involved in drug metabolism
- Assess the effects of these polymorphisms on metabolism of antibiotics

Genotyping results

| Cell Line | CYP2D6 | CYP2C9 | CYP3A4 | CYP2C19 | CYP3A5 | GSTM1 | GSTP1 | GSTT1 | NAT2 |
|-----------|--------|--------|--------|---------|--------|-------|-------|-------|-------|
| Huh-7D12 | Green | Green | Green | Green | Red | Green | Green | Red | Green |
| HepG2 | Green | Green | Green | Red | Red | Green | Green | Green | Red |
| SK-Hep-1 | Red | Green | Green | Green | Red | Green | Green | Green | Red |
| PLC/PRF/5 | Green | Green | Green | Red | Red | Red | Green | Red | Green |
| Hep3B | Green | Green | Green | Red | Red | Red | Red | Green | Green |

Figure 1. Genotyping using Taqman genotyping assays and conventional PCR-RFLP techniques. Two boxes per enzyme, representing each allele present:

Green Box = wild type allele, Red box = Mutant allele.

CYP enzymes chosen for genotyping are most polymorphic p450 oxidases involved in phase I metabolism. GST (Glutathione-S-Transferases) and NAT2 enzymes are involved in phase II of drug metabolism.

Conclusions

- Human Hepatoma cell lines tested in this work show genetic variation in selected enzymes involved in drug metabolism, with every cell line carrying at least four mutant alleles (Figure 1).
- We have shown that these cell lines metabolise a clinically relevant dose¹ of the antibiotic, Amoxicillin (Figure 2).
- There are differences in the levels of metabolism between the genetically different cell lines. The HepG2 cell line shows significantly higher levels of metabolism of amoxicillin (Figure 2).
- Preissner et al² suggests CYP2C19 responsible for metabolism of amoxicillin in humans, however our data indicates that this may not be the case.
- Although Cytochrome p450 class has more than 50 enzymes, just six of them metabolise 90% of drugs. CYP3A4 being one the most significant and most prevalent in the liver. Thus we believe CYP3A4 or CYP3A5 are more likely candidates for the metabolism of amoxicillin.
- We are currently using **CRISPR Gene Editing technology**³ to perform individual gene knock outs of CYP3A4, CYP3A5 and CYP2C19 enzymes in the HepG2 cell line. This may allow us to identify the enzyme (s) responsible for the metabolism of amoxicillin.

Acknowledgements

Many thanks to Ben Lee at University of Exeter Medical School for assistance with Taqman genotyping assays.

References

1. Szultka, M et al., Biomed. Chromatogr. 2014; 28: 255-264
2. Preissner et al., J Drug Metab Toxicol 2012, 3:5
3. Charpentier & Doudna 2013 Nature 495, 50-51